Bioactive potential
and possible health
effects of edible
brown seaweeds

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Nissreen Abu-Ghannam*

Introduction

Algae are heterogeneous group of plants with a long fos-
sil history. Due to their low content in lipids, high concen-
tration in polysaccharides, natural richness in minerals, polyunsaturated fatty acids and vitamins as well as their content in bioactive molecules, marine algae are known to be a good source of healthy food. Unlike the land plants, these algae have no roots, leaves or vascular systems; how-
ever they nourish themselves through the process of osmo-
sis. Two major types of algae that have been identified are the microalgae which are found in both benthic and littoral habitats and also throughout the ocean waters as phyto-
plankton and the macroalgae or seaweeds which occupy the littoral zone. Seaweeds grow in the intertidal as well as in the sub-tidal area up to a certain depth where very lit-
tle photosynthetic light is available. Seaweeds are classified into green algae (chlorophyta), brown algae (phaeophyta) and red algae (rhodophyta) on the basis of chemical composi-
tion. The color in case of green seaweeds is due to the presence of chlorophyll a and b in the same proportions as the 'higher' plants; beta-carotene (a yellow pigment) and various characteristic xanthophylls (yellowish or brownish pigments). The dominance of the xanthophyll pigment, fucoxanthin, is responsible for the color of brown seaweeds. This compound masks the other pigments such as Chlorophyll a and c and other xanthophylls. Phycoery-
thrin and phycocyanin mask the pigments such as Chloro-
phyll a and beta-carotene and are responsible for the

color of red seaweeds. Seaweeds are considered as a source of bioactive compounds as they are able to produce a great variety of secondary metabolites characterized by a broad spectrum of biological activities. They are an excellent source of vitamins such as A, B6, B12, C, D and E, ribofla-
vin, niacin, pantothenic acid and folic acid as well as min-
erals such as Ca, P, Na, K (Dharmananda, 2002). The fat content of seaweeds accounts for 1–6 g/100 g dry weight with some brown varieties, such as Hizikia sp. and Arame, having a fat content as low as 0.7–0.9 g/100 g dry weight (Kolb, Vallorani, & Stocchi, 1999). The red and the green species are rich in carbohydrates whereas the brown seaweeds are rich in soluble fiber and iodine. The highest iodine content is found in brown algae, with dry kelp (Laminaria) ranging from 1500 to 8000 ppm and dry rockweed (Fucus) from 500 to 1000 ppm (Dharmananda,
2002). Although seaweeds are exposed to the adverse envi-
ronmental conditions such as light and high oxygen con-
centrations that lead to the formation of free radicals, and other strong oxidizing agents, they do not have any serious photodynamic damage in vivo. Thus, it can be said that sea-
weds are able to generate the necessary compounds to pro-
tect themselves from external factors such as pollution, stress and UV radiation. This fact suggests that marine al-
gae, like photosynthesizing plants, have anti-oxidative
mechanisms and compounds which act as antioxidant agents. At the same time, several species of seaweeds have also been found to produce or contain polysaccharides, glycoproteins or other secondary metabolites with antimicrobial (Cox et al., 2010; Gupta, Rajauria, & Abu-Ghannam, 2010a), antitumoral (Koyanagi, Tanigawa, Nakagawa, Soeda, & Shimeno, 2003; Zubia et al., 2009) or anti-viral activity (Artan et al., 2008; Hemmingson, Falshaw, Furneaux, & Thompson, 2006; Zhu, Chiu, Ooi, Chan, & Angjr, 2004; 2003). Among all the three types highest phytochemical content have been reported from brown seaweeds (Seafoodplus, 2008). Thus, this review will mainly focus on the bioactive compounds present in the brown seaweeds. Recent developments in the isolation of compounds and characterization of the types of bioactive compounds from brown seaweeds will also be discussed. Focus is placed on the main classes of compounds that could be of medicinal and pharmaceutical value. The health benefits from the consumption of edible seaweeds and their role in nutrition is also explained.

Important metabolites from seaweeds
The division Phaeophyta consists of 13 orders according to the classification of Bold and Wynne (1985). However, only three orders namely Laminariales, Fucales and Dictyotales have been extensively researched for their phytochemicals. The most studied species of these orders are Laminaria, Ecklonia, Undaria, Himanthalia and Dictyota. In addition to being rich in polysaccharide, other important categories of metabolites found in brown seaweeds include polyphloroglucinol phenolic compounds (Ahn et al., 2004; Chandini, Ganesan, Suresh, & Bhaskar, 2008), non-polar, non-polyphenolic secondary metabolites such as terpenes (Siamopoulou et al., 2004), carotenoids such as fucoxanthin, volatile halogenated organic compounds (VHOCS) (Borchardt et al., 2001) and oxylipins (Kupper et al., 2006; Rorrer et al., 1995). This review will mainly focus on polysaccharides, polyphenolic compounds and terpenes in brown seaweeds.

Polysaccharides
Polysaccharides are a class of macromolecules which are increasingly gaining attention in the biochemical and medical areas due to their immunomodulatory and anti-cancer effects. These are present primarily in the cell walls and the composition varies according to season, age, species and geographic location. In addition to acting as a food reserve they also provide strength and flexibility to the plant to withstand wave action and maintain ionic equilibrium in the cell. The regularity of their structures also promotes interaction with external ions and inter-chain hydrogen bonding (e.g., gelation). Brown seaweeds are known to produce different polysaccharides, like alginates, fucoidans, and laminarans. Laminarans and fucoidans are the main water-soluble polysaccharides of brown algae whereas high-molecular mass algicic acids are alkali-soluble polysaccharides.

Cellulose microfibrils in cell wall of brown algae are embedded in an amorphous matrix of acid polysaccharide linked to each other by proteins. Brown algae have two kinds of acid polysaccharides present in the extracellular matrix: sulfated fucans and alginic acid. Fucans, (Fig. 1a), can be classified into three major groups: fucoidans, xylofucoglycuronans and glycorunogalactofucans. Fucoidan is a branched polysaccharide sulfate ester with 1-fucose 4-sulfate building blocks as the major

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**Fig. 1.** Structural unit of polysaccharides from brown algae (a) fucoidan; (b) laminaran.

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**Nomenclature**

<table>
<thead>
<tr>
<th>Symbol</th>
<th>Meaning</th>
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<tbody>
<tr>
<td>μM</td>
<td>Micro molar</td>
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<tr>
<td>d.w.</td>
<td>Dry weight</td>
</tr>
<tr>
<td>DPHC</td>
<td>diplorehydroxyxycarmalol</td>
</tr>
<tr>
<td>EC₅₀</td>
<td>Effective concentration of samples at which 50% effect is seen</td>
</tr>
<tr>
<td>GIT</td>
<td>Gastro intestinal tract</td>
</tr>
<tr>
<td>HCMV</td>
<td>Human cytomegalovirus</td>
</tr>
<tr>
<td>HIV</td>
<td>Human immunodeficiency virus</td>
</tr>
<tr>
<td>HSV</td>
<td>Herpes Simplex Virus</td>
</tr>
<tr>
<td>IC₅₀</td>
<td>Concentration at which 50% inhibition is achieved</td>
</tr>
<tr>
<td>kDa</td>
<td>Kilo Dalton</td>
</tr>
<tr>
<td>ppm</td>
<td>Parts per million</td>
</tr>
<tr>
<td>RT</td>
<td>Reverse transcriptase</td>
</tr>
<tr>
<td>SVHV</td>
<td>Sargassum vulgare high viscosity</td>
</tr>
<tr>
<td>SVLV</td>
<td>Sargassum vulgare low viscosity</td>
</tr>
<tr>
<td>UV</td>
<td>Ultra violet</td>
</tr>
<tr>
<td>VHOC</td>
<td>Volatile halogenated organic compounds</td>
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component. They are predominantly $\alpha(1\rightarrow2)$-linked with branching or a sulfate ester group at C$_3$ and is composed of fucose, uronic acids, galactose, xylose and sulfated fucose. The molecular weights reported for fucoidans vary in the range of approximately 100 kDa (Patankar, Oehninger, Barnett, Williams, & Clark, 1993) to 1600 kDa (Rupérez, Ahrazem, & Leal, 2002). Fucoidan is soluble in water and in acid solution (Rupérez et al., 2002). Acid hydrolysis of fucoidan yields various amounts of $\alpha$-xylose, $\alpha$-galactose, and uronic acid. Algal fucoidans are mainly found in Fucales and Laminariales, but are also present in Chordariales, Dictyoptales, Dictyosiphonales, Ectocarpales, and Sctyosiphonales. In fact, this kind of sulfated polysaccharide has been discovered in all the brown algae investigated so far, but seems to be absent in green algae, red algae, as well as in freshwater algae and terrestrial plants (Shanmugam & Mody, 2000). Xylofucoglycuranos or ascosphyllans consist of a polyuronide backbone, mainly poly-$\beta$-$$(1,4)$-$\beta$-mannuronic acid branched with 3-O-$\alpha$-xylosyl-1-fucose-4-sulfate or occasionally uronic acid. Glycuronogalactofucans are composed of linear chains of $(1,4)$-$\beta$-galactose branched at C$_3$ with L-fucosyl-3-sulfate or occasionally uronic acid (Jiménez-Escrig & Sánchez-Muniz, 2000).

Laminaran (or laminarin) was first discovered in Laminaria species and appears to be the food reserve of all brown algae. The major sugar of Laminaria species is laminaran whose structure and composition vary according to algae species. Laminaran is a water-soluble polysaccharide containing 20-25 glucose units which are composed of $(1,3)$-$\beta$-$\alpha$-glucan with $\beta(1,6)$ branching (Nelson & Lewis, 1974) (Fig. 1b). There are two types of laminaran chains (M or G), which differ in their reducing end. M chains end with a mannitol residue whereas G chains end with a glucose residue. Laminaran’s molecular weight is approximately 5000 Da depending on the degree of polymerization. Most laminarans form complex structures that are stabilized by inter-chain hydrogen bonds and are therefore resistant to hydrolysis in the upper gastro-intestinal tract (GIT) and are considered as dietary fibers (Neyrinck, Mouson, & Delzenne, 2007). The structure and the biological activities of laminaran and galactofucan are thought to be influenced by environmental factors, such as water temperature, nutritive salt, salinity, waves, sea current and depth of immersion. In addition to the role of laminarins as prebiotics and dietary fibers they have also been reported to possess antibacterial and anti-tumor activities.

Alginate acid or alginate is the common name given to a family of linear polysaccharides containing $1,4$-linked $\beta$-$\alpha$-mannuronic and $\alpha$-$\gamma$-guluronic acid (Fig. 2) residues arranged in a non-regular, block wise order along the chain (Andrade et al., 2004). Alginate produced by brown seaweed, especially in the form of sodium and calcium alginate, is widely used in the food and pharmaceutical industries due to their ability to chelate metal ions and to form highly viscous solutions.

**Fig. 2.** Monomeric compounds present in Alginic acid. (a) $\beta$-$\alpha$-mannuronic acid; (b) $\alpha$-$\gamma$-guluronic acid (Davis, Volesky, & Mucci, 2003).

Sulfated polysaccharides from marine algae have been described as possessing diverse biological activities with potential medicinal value, such as anti-coagulant, anti-tumor, anti-viral and anti-oxidant (Koyanagi et al., 2003; Ponce, Pujol, Damonte, Flores, & Stoerz, 2003; Shanmugam & Mody, 2000; Wijesekara, Pangestuti, & Kim, 2011 (and references therein)).

Other metabolites from seaweeds

Phlorotanins (Fig. 3) are tannin derivatives which are composed of phloroglucinol-based phenolics (1,3,5-trihydroxybenzene) and are synthesized via the acetate-malonate pathway. They are stored in special vesicles (physodes) and are thought to be the defense compounds in brown seaweeds. The concentration of phlorotanins in brown algae is reported to be highly variable among different taxa of brown seaweeds as well as among different geographical areas. Concentrations are reported to be higher in fucoid species and those obtained from the Atlantic and the temperate Pacific as compared to those obtained from the tropical Pacific (Targett & Arnold, 1998). Phlorotanins have secondary functions as defensive compounds and primary roles in cell-wall construction (Arnold & Targett, 2003).

Diterpenes (Fig. 4) are non-volatile halogenated compounds with different carbon structure including xenicane, dolabellane and prenylated guaiane skeletons (Blunt et al., 2009). Brown algae belonging to the genus Dictyota are a rich source of diterpenes. Dictyodial, dictyol C and dictyol H, which are typical algal terpenes, have been previously isolated from different species of Dictyota (Manzo et al., 2009). These secondary metabolites deter feeding by marine herbivores.

Volatile halogenated compounds such as bromophenols are common marine secondary metabolites, arising largely from the propensity of the phenol moiety to undergo electrophilic bromination. Bromophenols have been isolated from taxonomically diverse marine algae, for example, the brown algae *Fucus vesiculosus* and *Leathesia nana* (Xu et al., 2004a; 2004b). These compounds have been reported to act as a natural defense mechanism to prevent biofouling on the surface of Laminaria digitata by deactivation of acylated homoserine lactones (Borchardt et al., 2001). The presence of halogen substituent is unique for marine metabolites while it is rare for compounds obtained from terrestrial sources (Venkateswarlu, Panchagnula, Gottumukkala, & Subbaraju, 2007). The natural function of these compounds in seawater is uncertain, but it is often
suggested or assumed that they function as antimicrobial compounds or grazing deterrents.

Many marine macroalgae produce oxylipins, some of them belong to the prostaglandin and leukotriene series and share striking similarities with the products of cyclooxygenases and lipo-oxygenases in mammals. These oxylipins have been shown to play a role in chemical attraction and defense mechanisms. The formation of oxylipins in *L. digitata* was up-regulated in sporophytes challenged with lipopolysaccharides which may function as pathogen-associated molecular patterns (Kupper *et al.*, 2006). Ritter *et al.* (2008) reported that copper-induced

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**Fig. 3.** Chemical structures of different types of phlorotannins (Heo *et al.*, 2009; 2010).

**Fig. 4.** Structure of Diterpenes isolated from brown algae (Manzo *et al.*, 2009) (a) Dictyodial; (b) Dictyol C; (c) Dicytol H.
stress in *L. digitata* encouraged the accumulation of a number of complex oxylipins, which were thought to trigger protective mechanisms. Chemical attraction between female and male brown algal gametes is mediated by pheromones, such as hormonisirene and fucoserratene, which are hydrocarbons, thought to be down-stream products of a lipo-oxygenase pathway (Pohnert & Boland, 2002).

Fucoxanthin is the major biofunctional pigment present in brown seaweeds and is one of the most abundant carotenoids found in nature. It has a molecular structure consisting of an unusual allenic bond and a 5,6-monoepoxide. Fucoxanthin has been reported to have anti-oxidant and anti-tumor properties. Recently, it has been claimed that fucoxanthin can help in increasing the metabolism thereby controlling the weight gain in animal models (Maeda, Hosokawa, Sashima, & Miyashita, 2007). Heo, Yoon *et al.* (2010) studied the anti-inflammatory effect of fucoxanthin isolated from brown algae via inhibitory effect of nitric oxide production in lipopolysaccharide-induced RAW 264.7 macrophage cells.

Various methods have been used for the extraction and release of the bioactive compounds from seaweeds of which the use of organic solvents is most common. However, focus is now shifting to the use of green technologies such as enzyme assisted extraction (Heo, Park, Lee, & Yoon, 2006 and references therein) and various methods have been used for the extraction and release of the bioactive compounds from seaweeds of which the use of organic solvents is most common. However, focus is now shifting to the use of green technologies such as enzyme assisted extraction (Heo, Park, Lee, & Yoon, 2005; sub- and super-critical fluid for the extraction (Plaza, Cifuentes, & Ibáñez, 2008 and references therein; Herrero, Cifuentes, & Ibáñez, 2006 and references therein) of bioactive compounds.

### Bioactive properties of compounds from seaweeds

**Polysaccharides: anti-tumor, anti-viral, anti-coagulant**

Researchers have observed the effect of polysaccharides in biological systems as anti-coagulant, anti-tumor and anti-inflammatory agents (Table 1) and, which has led to the search for new compounds in the last few decades. Generally, the biological activity of polysaccharides from marine algae is related to the molecular size, type of sugar, sulfate content, type of linkage and molecular geometry which are known to have a role in their activities (Zhu *et al.*, 2004). Besides their well attested anti-coagulant and anti-thrombotic activity, they act on the inflammation and immune systems, have anti-proliferative and anti-adhesive effect on cells, protect cells from viral infection, and can interfere with mechanisms involved in fertilization.

**Anti-tumor properties**

Polysaccharides have shown good immunomodulatory properties associated with anti-tumor effects and thus search for these compounds in gaining attention. A role of sulfated polysaccharides from algae as anti-neoplastic agent has been suggested. Several investigations have reported that sulfated polysaccharides have anti-proliferative activity in cancer cell lines *in vitro*, as well as inhibitory activity against tumors growing in mice (de Souza, Marques *et al.*, 2007). Increasing the number of sulfate groups in the fucoidan molecule has been shown to affect the anti-tumor and anti-angiogenic activity (Koyanagi *et al.*, 2003).

### Table 1. Different algal bioactive compounds with possible effect on human health.

<table>
<thead>
<tr>
<th>Bioactive compounds</th>
<th>Specific compound</th>
<th>Possible health effect</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>F. evanescens</em></td>
<td>Fucoidan</td>
<td>Anti-tumor and Anti-metastatic</td>
<td>Alekseyenko <em>et al.</em>, 2007</td>
</tr>
<tr>
<td><em>F. vesiculosus</em></td>
<td>Fucan</td>
<td>Inhibitor of avian RT; Antithrombin</td>
<td>Queiroz <em>et al.</em>, 2008; Mourão, 2004</td>
</tr>
<tr>
<td><em>A. utricularis</em></td>
<td>Fucoidan</td>
<td>Inhibitory against HSV 1 and 2</td>
<td>Ponce <em>et al.</em>, 2003</td>
</tr>
<tr>
<td><em>L. japonica</em></td>
<td>Laminarin</td>
<td>Anti-apoptotic</td>
<td>Kim <em>et al.</em>, 2006</td>
</tr>
<tr>
<td><em>U. pinnatifida</em></td>
<td>sulfated polysac.</td>
<td>Anti-viral</td>
<td>Hemmingston <em>et al.</em>, 2006</td>
</tr>
<tr>
<td><em>E. cava</em></td>
<td>Phlorotannin</td>
<td>Whitening effect</td>
<td>Heo <em>et al.</em>, 2009</td>
</tr>
<tr>
<td><em>E. arborescens</em></td>
<td>Phlorotannin</td>
<td>Anti-allergy</td>
<td>Sugura <em>et al.</em>, 2007</td>
</tr>
<tr>
<td><em>I. okamure</em></td>
<td>Phlorotannin</td>
<td>Whitening effect; Anti-diabetic</td>
<td>Heo <em>et al.</em>, 2009, 2010</td>
</tr>
<tr>
<td><em>E. cava</em></td>
<td>Phlorotannin</td>
<td>Inhibitor of HIV-1 RT</td>
<td>Artan <em>et al.</em>, 2008; Ahn <em>et al.</em>, 2004</td>
</tr>
<tr>
<td><em>E. cava</em></td>
<td>Phlorotannin</td>
<td>Anti-cancer</td>
<td>Kong <em>et al.</em>, 2009</td>
</tr>
<tr>
<td><em>Pelvetia siliculosus</em></td>
<td>Phlorotannin</td>
<td>Anti-diabetic</td>
<td>Lee <em>et al.</em>, 2004</td>
</tr>
<tr>
<td><em>Ecklonia kurome</em></td>
<td>Phlorotannin</td>
<td>Algicidal</td>
<td>Nagayama <em>et al.</em>, 2003</td>
</tr>
<tr>
<td><em>Sargassum vulgare</em></td>
<td>Alginic acid</td>
<td>Antitumor</td>
<td>de Souza, Torres <em>et al.</em>, 2007</td>
</tr>
<tr>
<td><em>D. menstruus</em></td>
<td>Diterpenes</td>
<td>Anti-retroviral</td>
<td>Pereira <em>et al.</em>, 2004</td>
</tr>
<tr>
<td><em>D. sp.</em></td>
<td>Diterpenes</td>
<td>Cytotoxic</td>
<td>Jongarammuang &amp; Kongkam, 2007</td>
</tr>
<tr>
<td><em>D. platii</em></td>
<td>Diterpenes</td>
<td>Inhibitory against HSV-1; decrease the content of HSV-1 early proteins</td>
<td>Abrantes <em>et al.</em>, 2010</td>
</tr>
</tbody>
</table>
Dias et al. (2008) isolated a polysaccharide called as Sarg from the brown seaweed *Sargassum stenophyllum*, collected from Santa Catarina State, Brazil. The polysaccharide, Sarg, was studied for its anti-vasculogenic effects both in vivo and in vitro assays, as well as for its capacity to modify embryonic morphogenetic processes endogenously regulated by bFGF, a well-known angiogenic stimulator. Sarg could effectively inhibit vasculogenesis as well as developmental angiogenesis in chick embryos and could trigger concomitantly with vasculogenesis a specific change in the morphogenetic pattern.

Aisa et al. (2005) reported that fucoidan from *F. vesiculosus* inhibited the proliferation and induced apoptosis in human lymphoma HS-Sultan cell lines. They reported the fucoidan-induced apoptosis through a mitochondrial pathway as the mitochondrial potential in HS-Sultan cells was decreased 24 h after treatment with fucoidan.

Alekseyenko et al. (2007) studied the anti-tumor and anti-metastatic activities of fucoidan, isolated from *Fucus evanescens* present in Okhotsk sea, Russia in C57Bl/6 mice with transplanted Lewis lung adenocarcinoma. Fucoidan in a dose of 10 mg/kg and 25 mg/kg potentiated the anti-metastatic and anti-tumor activities of cyclophosphamide, respectively.

Kim, Kim, Kim, Lee, and Lee (2006) investigated the anti-apoptotic activity of laminaran polysaccharides isolated from the *Laminaria japonica*. The authors carried out a detailed pharmacological investigation on the laminaran polysaccharides and reported that it suppressed mouse thymocyte apoptosis and at the same time significantly induced the upregulation of 33 immunomodulatory genes from a total of 7410 genes which were examined using a cDNA microarray.

Alginites from brown seaweeds have also been reported to possess anti-tumor activity. de Sousa, Torres et al. (2007) investigated the in vivo anti-tumor activity of two alginites (*Sargassum vulgare* high viscosity (SVHV) and *S. vulgare* low viscosity (SVLV)) with different viscosity extracted from brown seaweed *S. vulgare* C Agardh, present in the Atlantic coast of Brazil, against Sarcoma 180 cells transplanted in mice. Both alginites could inhibit the growth of Sarcoma 180. The histopathological analysis of liver and kidney showed that both organs were affected by SVHV and SVLV treatment. However, only SVLV led to acute tubular necrosis. Alginites caused the enlargement of the white pulp of the spleen of treated animals, suggesting that the observed anti-tumor activity could be related to alginites immunomodulatory properties.

**Anti-viral property**

The anti-viral polysaccharides should have very low cytotoxicity toward mammalian cells if it is to be used for medicinal purposes and most of the algal polysaccharides have this attribute. Fucoidan has anti-viral properties toward viruses such as HIV and human cytomegalovirus (HCMV). Ponce et al. (2003) reported the presence of two different types of fucoidans, galactofuran and uronofucoidan, in the seaweeds *Adenocystis utricularis* collected from the shores near Comodoro Rivadavia, Argentina. The galactofuran showed a high inhibitory activity against herpes simplex virus (HSV) 1 and 2, with no cytoxicity whereas uronofucoidans had no anti-viral activity. The extraction of a polysaccharides fraction from aqueous extract of *Sargassum patens*, collected from Hong Kong coastal waters, has also been reported to be highly potent against HSV-1 and HSV-2 with an IC50 value as low as 25 µg/ml and 12.5 µg/ml, respectively. The polysaccharide had low levels of cytotoxicity toward mammalian cells (Zhu, Ooi, Chan, & Ang Jr., 2003). However, the characterization of this fraction has still not been done.

Chen, Lim, Sohn, Choi, and Han (2009) studied the inhibitory effects of fucoidan, isolated from *Undaria pinnatifida* collected from north east coast of Korea, on the growth of *Plasmodium falciparum* parasites in vitro and on *Plasmodium berghei*-infected mice in vivo. Fucoidan significantly inhibited the invasion of erythrocytes by *P. falciparum* merozoites, and its IC50 was found to be similar to those for the chloroquine-sensitive *P. falciparum* 3D7 strain and the chloroquine-resistant K1 strain. Queiroz et al. (2008) assessed the activity of fucans isolated from *F. vesiculosus* (from the coast of Natal, Brazil) as inhibitors of HIV from reverse transcriptase (RT). These fucans had a pronounced inhibitory effect in vitro on the avian-RT at a concentration of 0.5–1.0 µg/mL. The alginic acid (1.0 mg/mL) inhibited the RT activity by 51.1% using activated DNA. The authors attributed the inhibitory to the fucans to the presence of sulfate groups as desulphation resulted in the loss of this effect. Furthermore it was suggested that fucan activity was not only dependent on the ionic changes but also on the sugar rings that act to spatially orientate the charges in a configuration that recognizes the enzyme, thus determining the specificity of the binding (Queiroz et al., 2008). Hemmingson et al. (2006) studied the anti-viral activity of a galactofucan sulfate extract from *U. pinnatifida* collected from east coast of Tasmania, Australia. It was found to be a potent inhibitor of the herpes viruses HSV-1, HSV-2 and HCMV, with IC50 values of 1.1, 0.2 and 0.5 µg/mL, respectively.

**Anti-coagulant property**

Anti-coagulant property is another widely studied property of sulfated polysaccharides. Anti-coagulant activity of sulfated polysaccharides has been identified from several brown seaweeds such as *Padina gymnospora* (Silva et al., 2005), *Dictyota menstrualis* (Albuquerque et al., 2004) and *F. vesiculosus* (Mourão, 2004).

Yoon, Pyun, Hwang, and Mourão (2007) isolated an acidic polysaccharide from *Laminaria cichorioides* collected from the coast of Korea which was shown to have a potent anti-coagulant activity mainly mediated by thrombin inhibition by heparin cofactor II. Studies using a sulfated fucan from *F. vesiculosus* suggested that the
antithrombin activity is mediated mainly by heparin cofactor II, with a minor contribution of antithrombin (Mourão, 2004).

**Diterpenes**

Diterpenes have been reported to have cytotoxic, antiviral and algicidal activities (Table 1). Several types of diterpenoids, such as dolabellanes, hydroaizulenoids, xenocanes and so-called extended sesquiterpenoids, have been found to be the main secondary metabolites of the species belonging to the Dictyotaceae family.

**Anti-tumor property**

Two diterpenes, 4,18-dihydroxydictyolactone 132 and 8a,11 dihydroxyphachydictyl A 133, were isolated from a *Dictyota* sp. collected from Bangsaen Beach, Thailand (Jongaramruong & Kongam, 2007). In bioassays, 4,18-dihydroxydictyolactone was strongly cytotoxic (NCI-H187) (Jongaramruong & Kongam, 2007). Awad, Selim, Metawe, and Matloub (2008) isolated 18,19 epoxyxenic-19-methoxy-18-hydroxy-4-acetoxy-6,9,13-triene and 18,19 epoxyxenic-19,18 dimethoxy-4-hydroxy-6,9,13-triene from methanol extracts of *Padina pavonia* collected from the Red Sea at Harghada, Egypt. The isolated compounds showed anti-tumor activities against lung carcinoma (H460) and liver carcinoma (HepG2) human cell lines (in vitro). Zabia et al. (2009) assessed the anti-oxidant and anti-tumoral activities of crude extracts from 10 phaeophyta species from Brittany coasts. Anti-tumoral activities were determined by a cytotoxic assay with three different tumoral cells lines (Daudi, Jurkat and K562). Five species exhibited strong cytotoxic activities against all tumoral cells. The cytotoxic effect was attributed to the high level of diterpenes compounds in the Sargassaceae species used in the study.

**Anti-viral property**

Soares et al. (2007) isolated meroditerpenoids atomaric acid, epitaenolid and the peroxyxactone of 5α-desmethyl-5α-acetylatomaric acid from Brazilian seaweed *Stypodium zonale*. These compounds showed strong anti-HSV-1 activity in vitro but none could inhibit the transcriptase reverse enzyme of HIV-1. Pereira et al. (2004) studied the effect of two diterpenes (6R)-6-hydroxydictohotama-3,14-diene-1,17-dial, named Da-1, and (6R)-6-acetoxy-dichotoma-3,14-diene-1,17-dial, named AcDa-1) isolated from Brazilian seaweed, *Dictyota menstrualis*, on HIV-1 replication. The compounds were reported to have an effect on an early step of the virus replicative cycle or during virus adsorption/penetration. The isolated compounds were shown to inhibit the RNA-dependent DNA-polymerase activity of HIV-1 RT in a dose-dependent manner with an EC_{50} of 40 μM and 70 μM. However, the diterpenes were not as strong as the well-known non-nucleoside inhibitor of the HIV-1 RT nevirapine (EC_{50} 40 nM). Siamopoulou et al. (2004) also reported anti-viral activity of diterpenes isolated from *D. dichotoma* collected from the coasts of Saronikos gulf in Athens and *D. linearis* from the south coasts of Chios Island. The isolated metabolites did not exhibit significant anti-viral activity against against Poliomyelitis virus I and HSV-1 in concentrations lower than their maximal non-toxic dose. Abrantes et al. (2010) reported the inhibition of HSV-1 infection in vero cells with diterpenes 8,10,18-trihydroxy-2,6-dolabeladiene and (6-R)-6-hydroxydictohotama-4,14-diene-1,17-dial, isolated from the Brazilian marine algae *Dictyota pfaffii* and *D. menstrualis*, respectively. The compounds inhibited HSV-1 replication in a dose-dependent manner, resulting in EC_{50} values of 5.10 and 5.90 μM, respectively, for a multiplicity of infection of 5. In addition, the tested molecules could decrease the contents of some HSV-1 early proteins, such as UL-8, RL-1, UL-12, UL-30 and UL-9.

**Phlorotannins**

Phlorotannins have been clarified to exhibit anti-diabetic (Lee, Shin, Kim, & Lee, 2004), anti-oxidation (Ahn et al., 2007), anti-cancer (Kong, Kim, Yoon, & Kim, 2009; Yang, Zeng, Dong, Liu, & Li, 2010), and anti-HIV (Ahn et al., 2004) (Table 1) activities.

**Anti-oxidant property**

Heo, Ko et al. (2009) isolated three kinds of phlorotannins from *Ecklonia cava* collected from the coast of Jeju Island, Korea and studied their inhibitory effect on melanogenesis as well as their protective effect against oxidative stress induced by UV-B radiation. They reported that the phlorotannin, dieckol, has potential whitening effects and prominent protective effects on UV-B radiation-induced cell damages. Dieckol showed 88.9% tyrosinase inhibitory activity even at 50 μM, and the values were higher than that of commercial whitening agent, kojic acid. Heo, Ko et al. (2010) also isolated diphlorethohydroxycarmalol (DPHC) from *Ishige okamurae* extracts. DPHC demonstrated strong protective properties against UV-B radiation via damaged DNA tail length and morphological changes in fibroblast, thus showing that the compound has a potential whitening effect and can have potential use in the pharmaceutical and cosmetic industry.

**Anti-allergic property**

The anti-allergic properties of several phlorotannins isolated from seaweeds have been studied on leukemia cell lines in vitro. Sugiiura et al. (2007) isolated a phlorotannin, phlorofucofuroeckol-B, from *Eisenia arborea* collected from the Mugizaki coast in Mie prefecture, Japan. The compound was reported to have anti-allergic properties. The isolation was guided by the inhibitory effect of the collected fractions from the extract on histamine release (IC_{50} 7.8 μM) from rat basophile leukemia (RBL-2H3) cells in a concentration-dependent manner. Le, Li, Qian, Kim, and Kim (2009) isolated two main bioactive phlorotannin derivatives together with phloroglucinol and dieckol having
anti-allergy activity from crude extracts of Korean seaweed *Ecklonia cava*. The anti-allergic activity of these derivatives was assessed by histamine release assay on human basophilic leukemia (KU812) and rat basophilic leukemia (RBL-2H3) cultured cell lines, respectively. Strong inhibitory effect was shown by dieckol and one phlorotannin derivative.

**Anti-diabetic property**

In *vivo* testing of fucosterol in streptozotocin-induced diabetic rats, isolated from the brown alga *Pelvetia siliculososa*, demonstrated that it is the main anti-diabetic principle (Lee et al., 2004). Fucosterol caused a significant decrease in serum glucose concentrations, and exhibited an inhibition of sorbitol accumulations in the lenses of rats (Lee et al., 2004). Heo, Hwang et al. (2009) reported that diphloethyloxyacarmalol (DHC) isolated from *I. okamurae* collected along the Coast of Jeju Island, Korea might be a potent inhibitor of *α*-glucosidase and *α*-amylase. The IC\textsubscript{50} values of DPHC against *α*-glucosidase and *α*-amylase were 0.16 and 0.53 mM, respectively, which evidenced the higher activities than that of acarbose. Moreover, DPHC did not seem to exert any cytotoxic effect in human umbilical vein endothelial cells at various concentrations (from 0.49 to 3.91 mM). At the same time, the increase of postprandial blood glucose levels were significantly suppressed in the DPHC-administered group than those in the streptozotocin-induced diabetic or normal mice.

**Anti-viral property**

Tannins have been reported to show their HIV-1 inhibitory mode of action by inhibiting polymerase and ribonuclease activities of HIV-1 RT (Artan et al., 2008). Ahn et al. (2004) isolated four phlorotannin derivatives, eckol (1), 8,8'-dieckol (2), 8,4'-dieckol (3), and phlorofucofuranoeckol A (4) from *E. cava*. Among these, compounds 2 (IC\textsubscript{50}, 0.51 μM) and 3 (IC\textsubscript{50}, 5.3 μM) exhibited an inhibitory effect on HIV-1 RT. Both these compounds were able to inhibit the protease as well but RT was inhibited more potently than the protease. Another phlorotannin 6,6'-dieckol was isolated from *E. cava* and studied for its antiviral properties (Artan et al., 2008). The compound showed wild inhibition against HIV-1 induced syncytia formation (EC\textsubscript{50} 1.72 μM), lytic effects (EC\textsubscript{50} 1.23 μM), and viral p24 antigen production (EC\textsubscript{50} 1.26 μM), respectively in addition to inhibiting the activity of HIV-1 RT enzyme with EC\textsubscript{50} of 1.07 μM, as well as HIV-1 entry.

**Anti-tumor property**

Kong et al. (2009) isolated phloroglucinol derivatives, dioxinodehydroeckol (1) and 1-(3',5',6-trihydroxyphenoxy)-7-(2',4',6-trihydroxyphenoxy)-2,4,9-trihydroxydibenzoyl-1,4-dioxin (2), from *E. cava* and checked their ability to inhibit the proliferation of human breast cancer cells. Compound 1 exerted a higher anti-proliferative activity in human breast cancer cells, induced a significant proliferative inhibition and apoptosis in a dose-dependent manner on MCF-7 human cancer cells and also induced the increase in caspase (~3 and ~9) activity. Yang et al. (2010) studied the anti-proliferative activity of phlorotannins derived from brown algae *L. japonica* Aresch extracts collected from Quingdao, China on the human hepatocellular carcinoma cell (BEL-7402) and on murine leukemia cells (P388) by MTT assay. The inhibitory rate of phlorotannin extract on BEL-7402 and P388 cells was 30.20 ± 1.16% and 43.44 ± 1.86%, respectively, and IC\textsubscript{50} on P388 and BEL-7402 cells was 120 μg/mL and >200 μg/mL, respectively.

**Antibacterial and algicidal property**

In addition, bactericidal (Nagayama, Iwamura, Shibata, Hirayama, & Nakamura, 2002) and algicidal activity (Nagayama, Shibata, Fujimoto, Honjo, & Nakamura, 2003; Wang, Xiao, Wang, Zhou, & Tang, 2007) of phlorotannins has also been reported. Nagayama et al. (2003) reported phlorofucofuroeckol A, to have algicidal activity as strong as that of epigallocatechin gallate. Nagayama et al. (2002) found the bactericidal effect of the phlorotannins to be more pronounced than those of the catechins which was used as positive control. Wang, Xu, Bach, and MacAllister (2009) reported the bactericidal effects of phlorotannins isolated from *Ascophyllum nodosum* collected from Atlantic coastline of Nova Scotia, Canada against *E. coli* O157:H7. The marine phlorotannins were reported to be superior in activity as compared to terrestrial phlorotannins.

While all these studies show substantial evidence to suggest that seaweed phytochemicals have the potential to be used as nutraceuticals or in pharmaceutical industry, to date not much progress has been made on *in vivo* activity of these compounds isolated from seaweeds.

**Health benefit due to consumption of seaweed dietary fibers**

Being rich in polysaccharides which are not digested by intestinal enzymes makes seaweeds an important source of dietary fibers and can be considered as a source of prebiotics. A prebiotic is a compound which must be resistant to digestion in the upper GIT and therefore resistant to acid and enzymatic hydrolysis; must be a selective substrate for the growth of beneficial bacteria and finally, it must induce luminal or systemic effects that are beneficial to host health. These dietary fibers differ chemically and physico-chemically from that of the terrestrial species and may induce different fermentative patterns. The content of total dietary fiber in seaweeds ranges from 33 to 50 g/100 g d.w. (Rupérez & Saura-Calixto, 2001). Accordingly, the fiber content of seaweed varieties is higher than those found in most fruits and vegetables. The human consumption of algal fiber has been proven to be health-promoting and its benefits are well documented in the scientific literature. The consumption of this dietary fiber has been related to the following health-promoting effects:
(1) promotes the growth and protection of the beneficial intestinal flora, (2) reduces the overall glycemic response, (3) greatly increases stool volume and (4) reduces the risk of colon cancer. In addition to the presence of some of the components which have potential benefits for the human body, the presence of dietary fibers provides some technological advantages for the use of marine algae as ingredients in food products such as meat products. The presence of these prebiotics can also be used to support the growth of lactic acid bacteria using seaweed broth as a sole source of nutrition (Gupta, Abu-Ghannam, & Scanell, 2010b) and subsequently probiotics that can benefit human health. Thus, seaweeds have the potential to be used as a functional food ingredient or as a nutraceutical.

The capacity of the fibers to absorb and retain water (Rupérez & Saura-Calixto, 2001) helps in the utilization of seaweeds as texturing and bulking agents, particularly in the making of low calorie foods. At the same time, the high concentration of mineral elements in seaweeds can help to reduce the amount of added sodium chloride in meat processing. López-López et al. (2009) studied the influence of the addition of edible seaweeds Himanthalia elongata, U. pinnatifida and Porphyra umbilicatis collected from the Atlantic coast, on fatty acid composition, amino acid profile, protein score, mineral content and antioxidant capacity in low-salt meat emulsion model systems. Meat systems made with added seaweeds had lower (P < 0.05) sodium contents than control samples. The inclusion of H. elongata increased the sulfur amino acid score by 20%. The added seaweeds supplied the meat samples with soluble polyphenolic compounds, which increased the anti-oxidant capacity of the systems.

The prebiotic effect of seaweed polysaccharide was shown by its ability to resist digestion in the upper GIT in a study conducted by Deville, Damas, Forget, Dandrifosse, and Peulen (2004). They reported that laminarin remained intact following incubation in vitro with hydrochloric acid, human saliva and human gastric, pancreatic, small intestinal and colonic homogenates. Feeding trials have also been performed in laboratory animals to investigate the effects on animal health and growth performance. Guidel-Urbano and Goñi (2002) studied the influence of feeding two edible seaweeds, Porphyra and Undaria purchased from a local health store in Madrid, Spain, as a source of dietary fiber on dietary nutritive utilization in male adult Wistar rats. The addition of seaweed did not affect the gain in body weight of rats or food efficiency but the fresh and dry stool weights were higher in rats fed seaweeds than in the control group. Seaweed-fed animals showed significantly lower apparent digestibilities of protein and fat but absorbed nitrogen was more effectively used by animals. Evidence is also available on the prebiotic effect of seaweed polysaccharide on animal health. Kuda, Yano, Matsuda, and Nishizawa (2005) reported that dietary supplementation with 1% laminarin resulted in an increase in Bifidobacterium counts in the cecum of rats compared to a control diet, but there was no significant difference in Lactobacillus counts. Wang, Han, Hu, Li, and Yu (2006) reported a selective increase in the numbers of Bifidobacterium and Lactobacillus in both the cecum and faeces of rats which were fed diets supplemented with 2.5% alginate. The prebiotic effect was found to be greater than the control group which was fed on a diet containing prebiotic fructo-oligosaccharride. Deville, Gharbi, Dandrifosse, and Peulen (2007) noted that laminarin can influence the adherence and the translocation of bacteria across the epithelial wall and seems to be a modulator of the intestinal metabolism by its effects on mucus composition, intestinal pH and short-chain fatty acid production, especially butyrate. Neyrinck et al. (2007) demonstrated that dietary supplementation with laminarin protected against lipopolysaccharide-induced liver toxicity in a rodent model of systemic inflammation. They suggested that the immunomodulatory effects of dietary laminarin could be either due to a direct effect of laminarin on immune cells or due to an indirect effect via modulation of the intestinal microbiota. Maeda et al. (2007) studied the anti-diabetic and anti-obesity effect of dietary fucoxanthin and fish oil. They reported that dietary fucoxanthin decreases the blood glucose and plasma insulin concentration of KK-A' along with down-regulating tumor necrosis factor-α mRNA. Reports are also available on the effect of feeding of farm animals with whole seaweeds or seaweed polysaccharide. Lynch, Sweeney, Callan, O’Sullivan, and O’Doherty (2010) showed the prebiotic effect of feeding pigs with laminaran and fucoidan on intestinal fermentation and selected microflora. Feeding resulted in a reduction in intestinal Enterobacteria and an increase in Lactobacilli sp thus suggesting that feeding of seaweeds can act as a dietary means to improve gut health in pigs. Reilly et al. (2008) demonstrated the effect of dietary supplementation with extracts containing laminarin and fucoidan from different varieties of brown seaweeds, L. digitata and Laminaria hyperborea collected from Kerry, Ireland on gut morphology and intestinal microbial populations in pigs. The inclusion resulted in an inhibitory effect on the Enterobacteria, Lactobacilli and Bifidobacteria population within the caecum and colon of weaned pigs. O’Doherty, McDonnell, and Figat (2010) showed that feeding laminarin resulted in the reduction in faecal E. coli population and an increase in daily gain and gain to feed ratio to improve gut health in post weaning pigs. However, a combination of laminarin and fucoidan was reported to be more effective at reducing diarrhea post weaning. Dillon, Sweeney, Callan, and O’Doherty (2010) have also reported that the inclusion of a combination of laminarin and fucoidan extract derived from L. digitata, increased daily gain and gain to feed ratio of post weaned piglets. According to the authors this was mainly due to an increase in nutrient digestibility and decreased E. coli populations in the guts. Dierick, Oyyn, and De Smet (2010) studied the effect of feeding intact A. nodosum collected from Ireland on
the piglet gut flora (E. coli, lactobacilli, streptococci, total anaerobic count) and their metabolism. In vitro investigations, simulating in vivo conditions, revealed a statistically significant depressive effect of seaweed on piglet small intestinal and hindgut flora, especially on E. coli. Also the fermentative activity (lactic acid, volatile fatty acids) of the flora was lowered.

Conclusions

Seaweeds grow in abundance in coastal areas and are available all year round. This review attempted to examine the reports available on the compounds being isolated from seaweeds that may have anti-cancer, anti-tumor or antiviral activity. Many reports have been published about isolated compounds from algae with biological activity, demonstrating their ability to produce metabolites however a lot of research is needed before this vast untapped resource could be utilized for beneficial purposes. Thus, the investigation of new algal chemical compounds, a different source of natural products, can prove to be a promising area of pharmaceutical study. Moreover, substantial amount of research regarding the toxicity aspects also needs to be carried out before they could actually be used for clinical trials. The information available on the probiotic potential of seaweeds being fed to farm animals seems promising. However, the results from different studies are conflicting and more studies are needed in order to reach a consensus regarding their beneficial dietary effect. At the same time, they may also be a source of compounds which could be exploited for novel functional ingredients for human and animal health applications. Future work in the area of seaweed-derived bioactives should aim to examine the effects of purified compounds under in vivo conditions to understand their actual potential.

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References


Arnold, T. M., & Targett, N. M. (2003). To grow and defend: lack of fermentative activity (lactic acid, volatile fatty acids) of intestinal and hindgut flora, especially on free radical scavenging using ESR and H2O2- mediated DNA damage.


Worth it!


cystis utricularis*: extraction methods, antiviral activity and struc-


Rupérez, P., & Saura-Calixto, F. (2001). Dietary fibre and physico-
chemical properties of edible Spanish seaweeds. *European Food Research and Technology, 212*, 349–354.

Rupérez, P., Abrazeno, O., & Leal, J. A. (2002). Potential antioxidant ca-


tion and anticoagulant activity of a heterofucan from the brown seaweed *Padina gymnospora*. *Brazilian Journal of Medical and Biological Research, 38*, 523–533.


Wang, Y., Han, F., Hu, B., Li, J. B., & Yu, W. G. (2006). In vivo prebiotic properties of alginate oligosaccharides prepared through enzy-


